

BIOSYNTHESIS OF A VITAMIN K₂ BY CELL-FREE EXTRACTS
OF MYCOBACTERIUM PHLEI^{*}

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Very little is known of the de novo biosynthesis of vitamin K₂ by microorganisms. Cox and Gibson (1964) have described the incorporation of ¹⁴C-shikimic acid into vitamin K₂(35) in whole cells of E. coli. We have recently found that whole cells of M. phlei incorporate the radioactivity from L-methionine-¹⁴CH₃ into vitamin K₂(45)H^{MM} (Lederer, 1964; Azerad et al., 1965). We now report the incorporation of radioactivity from L-methionine-¹⁴CH₃ into vitamin K₂ (45)H by a cell-free supernatant obtained from M. phlei.

METHODS. M. phlei Legroux cells were grown in Sauton medium for 7-8 days. Twenty g. of washed cells were suspended in 20 ml of a KCl-tris buffer (1.15 % KCl in 0.05 M tris, pH 7.8) and were sonicated for 8 minutes at 0°C in a Raytheon 10 kc ultrasonicator. The supernatant obtained after centrifugation for 10 minutes at 10,000 x g was used for the incorporation experiments, the conditions being detailed in Table I. ATP and MgCl₂ were added to facilitate the formation of S-adenosyl-methionine; further additions of ATP were made at 1 and 2 hours during the 3-hour incorporation period. The reaction was stopped by precipitation of proteins with acetone (3 vol. /vol. of supernatant) and the subsequent filtrate was evaporated to dryness prior to extraction with

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^{MM} M. phlei contains a vitamin K₂(45)H, i. e., a 2-methyl 1,4-naphtoquinone with a C₄₅ isoprenoid side chain in position 3; one of the isoprenoid units is saturated (Gale et al., 1963); we have confirmed by mass spectrometry the molecular formula proposed by these authors (Lederer, 1964; mass spectrum by Dr. M. Barber, A. E. I., Manchester).

TABLE I

Aerobic incubation of *M. phlei* supernatant with L-methionine- $^{14}\text{CH}_3$ and L-tyrosine-U- ^{14}C for 3 hours at 30°C.

Composition of incubation medium ^x	Dose μcuries	Specific activity dpm/mg of vit. K ₂ (45)H
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L-methionine- $^{14}\text{CH}_3$ (sp. act. 9.3 mcuries/mmole)		
" + cell-free supernatant, 45 ml	45	12,250
" "	22.5	13,000
" "	22.5	11,500
" + boiled supernatant, 45 ml	45	<200
L-tyrosine-U- ^{14}C (sp. act. 72.6 mcuries/mmole) + ammonium acetate, 90 μmoles + DL-methionine ^{xx} , 90 μmoles		
" + cell-free supernatant, 45 ml	10	300
" + boiled supernatant, 45 ml	10	<300

^xAll incubation media contained: MgCl_2 , 4.5 mmole; ATP, 225 μmoles (+90 μmoles added at 1 and 2 hours during incubation) in KCl-tris (1.15 % in 0.05 M, pH 7.4); final volume: 8.5 ml.

^{xx}DL-methionine was dissolved in the KCl-tris buffer made slightly acidic with HCl.

hexane. The vitamin K₂(45)H in the hexane extract was purified by repeated thin layer chromatography on Kieselgel G (Merck) in the solvent system hexane-ethyl methyl ketone (97:3), until constant specific activity was reached. In addition, the purity of vitamin K₂(45)H was checked by UV spectrum, reversed phase thin layer chromatography (Kieselgel G plates treated with 10 % vaseline in CHCl_3 ; solvent, acetone-water (95:5), $R_f = 0.4$), and reversed phase ascending paper chromatography (Whatman No. 1 impregnated with vaseline 5 % in hexane; solvent, acetone-water (95:5), $R_f = 0.25$).

Vitamin K₂(45)H was determined quantitatively by reduction with NaBH_4 in buffered alcohol, pH 5 (Lester *et al.*, 1964) using

$E_{1\text{cm}}^{1\%}$ ox. (271 m μ)- $E_{1\text{cm}}^{1\%}$ red. (271 m μ) = 166; radioactivity was measured with a Packard 3003 Tricarb Scintillation Spectrophotometer.

RESULTS. Three successive purifications on thin layer plates of the hexane-extracted vitamin K₂ sufficed to obtain a product assessed pure by the criteria described in the preceeding section.

In three experiments as shown in Table I, an average of 12,250 dpm/mg of vitamin K₂(45)H were incorporated from L-methionine-¹⁴CH₃ during a 3-hour incubation period. That maximum incorporation was obtained under these conditions is indicated by the specific activities determined when L-methionine-¹⁴CH₃ was used at two concentrations. When the supernatant from M. phlei was boiled for 5 minutes, prior to its addition to the incubation system, radioactivity was not incorporated into vitamin K₂(45)H; preformed vitamin K was not destroyed during the boiling process.

L-tyrosine-U-¹⁴C, added to the cell-free supernatant system, was not incorporated into vitamin K₂(45)H (Table I); boiled supernatant also served as a control in this experiment.

DISCUSSION. In this paper, we describe a cell-free supernatant from M. phlei that incorporates radioactivity of L-methionine-¹⁴CH₃ into vitamin K₂(45)H. This finding confirms our previously reported results (Lederer, 1964; Azerad *et al.*, 1965) on the incorporation of radioactivity from L-methionine-¹⁴CH₃ into vitamin K₂(45)H by whole cells of M. phlei.

The 2-methyl group of ubiquinone 50 is derived from methionine, as shown by Rudney and Parson (1963) (see also Olson *et al.*, 1963). Work is in progress to locate the labelled carbon in the radioactive vitamin K obtained in the experiments described above and to investigate the biosynthesis of the naphthalene nucleus of this vitamin by means of our cell-free system. Tyrosine does not appear to be a precursor in the biosynthetic pathway to vitamin K₂(45)H in this system.

Addendum. After writing this paper, we have learned that Martius and Leuzinger (1964) have described the incorporation of the radioactivity

of $^{14}\text{CH}_3$ -methionine into vitamin $\text{K}_2(45)$ by whole cells of the anaerobe: Fusiformis nigrescens, starting from 1,4-naphtoquinone.

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